

Primer Design Ltd

R01045

Mycobacterium (all species)

Kit version: 2

Target region:
16S rRNA gene

genesig[®] Standard Kit

150 tests

For general laboratory and research use only

GENESIG

Kits by Primerdesign

Introduction to Mycobacterium (all species)

Mycobacteria are gram-positive, catalase positive, non-motile, non-spore forming rod-shaped bacteria. The bacteria are 0.2-0.6 μm wide and 1.0-10 μm long and can have various colony morphologies; rough or smooth and can appear white-orange-pink. Mycobacteria are primarily aerobic however there are some species which are microaerophilic.

Mycobacteria have thick hydrophobic cell walls consisting of four layers, the inside layer is formed from peptidoglycan and the outer three of lipids. Lipid layers confer resistance to acidic and alkaline environments and make the cell impermeable to basic dyes.

Mycobacterium tuberculosis is the most clinically important mycobacterium species as it causes tuberculosis in humans, *Mycobacterium africanum* causes tuberculosis in humans in central Africa and *Mycobacterium bovis* is able to cause tuberculosis in both cattle and humans. These three species are genetically similar and comprise the *Mycobacterium tuberculosis* complex (these are all obligate pathogens which require a mammalian host). Other pathogenic mycobacteria species are grouped into the *Mycobacterium avium* complex (MAC) and non-tuberculosis mycobacteria (NTM). Infections by these species particularly affect immunosuppressed individuals.

Most of the identified *Mycobacterium* species are opportunistic environmental pathogens (considered opportunistic saprophytes) found in soil and water. These species can be further divided into two groups, slow growing mycobacteria (for example MAC) and rapidly growing mycobacteria (formerly known as atypical mycobacteria).

Real-time PCR can be used for the rapid detection of *Mycobacteria* species.

Specificity

The genesig® Standard Kit for Mycobacterium (all species) is designed for the in vitro quantification of Mycobacterium genomes. The kit is designed to have a broad detection profile. Specifically, the primers will detect over 95% of sequences available on the NCBI database at the time of the most recent review.

This kit is predicted to cross react with Mycobacteriodes, Mycolicibacter, Mycolicibacterium and Mycolicibacillus, four genera recently separated from the Mycobacterium genus. The kit is also predicted to cross react with certain members of the Microbacterium, Gordonia, Rhodococcus, Nocardia and Corynebacterium generas.

The dynamics of genetic variation means that new sequence information may become available after the initial design. If you require further information or have a specific question about the detection profile of this kit, then please send an e-mail to techsupport@primerdesign.co.uk and our team will answer your question.

Kit contents

- **1x Mycobacterium spp_v2.0 primer/probe mix (150 reactions, BROWN)**
FAM labelled
- **1x Mycobacterium spp_v2.0 positive control template (RED)**
- **1x RNase/DNase free water (WHITE)**
for resuspension of primer/probe mixes
- **2x Template preparation buffer (YELLOW)**
for resuspension of positive control template and standard curve preparation

Reagents and equipment to be supplied by the user

Real-time PCR Instrument

Extraction kit

This kit is recommended for use with genesig® Easy DNA/RNA extraction kit or exsig®Mag. However, it is designed to work well with all processes that yield high-quality nucleic acid with minimal PCR inhibitors.

oasig® lyophilised or PrecisionPLUS® 2X qPCR Master Mix

This kit is intended for use with oasig® lyophilised or PrecisionPLUS® 2X qPCR Master Mix.

Pipettors and filter tips

Vortex and centrifuge

1.5 ml microtubes

qPCR plates or reaction tubes

Kit storage and stability

This kit is stable for shipping at ambient temperature but should be stored at -20°C upon arrival. Once the lyophilised components have been resuspended, they should not be exposed to temperatures above -20°C for longer than 30 minutes at a time and unnecessary repeated freeze/thawing should be avoided. The kit is stable for six months from the date of resuspension under these circumstances.

If a standard curve dilution series is prepared this can be stored frozen for an extended period. If you see any degradation in this serial dilution a fresh standard curve can be prepared from the positive control.

Primer Design Ltd does not recommend using the kit after the expiry date stated on the pack.

Suitable sample material

This kit can be used with all types of samples from various origins. Please ensure that the extracted nucleic acid samples are suitable in terms of purity, concentration, and DNA integrity.

Dynamic range of test

Under optimal PCR conditions the kit can achieve priming efficiencies between 90-110% and detect less than 100 copies of target template. If running a positive control standard curve for a quantitative result, and an efficiency of between 90% to 110% is not achieved, then the run should be repeated with a freshly prepared standard curve.

Principles of the test

Real-time PCR

A target specific primer and probe mix is provided, and this can be detected through the FAM channel.

The primer and probe mix provided exploits the so-called TaqMan[®] principle. During PCR amplification, forward and reverse primers hybridize to the target DNA. A fluorogenic probe is included in the same reaction mixture which consists of a DNA probe labelled with a 5'-dye and a 3'-quencher. During PCR amplification, the probe is cleaved, and the reporter dye and quencher are separated. The resulting increase in fluorescence can be detected on a range of qPCR platforms.

Positive control

For copy number determination and as a positive control for the PCR set up, the kit contains a positive control template. This can be used to generate a standard curve of target copy number/Cq value. Alternatively, the positive control can be used at a single dilution where full quantitative analysis of the samples is not required. Each time the kit is used, at least one positive control reaction must be included in the run. A positive result indicates that the primers and probes for detecting the target gene worked properly in that particular experimental scenario. If a negative result is obtained the test results are invalid and must be repeated. Care should be taken to ensure that the positive control does not contaminate any other kit component which would lead to false-positive results. This can be achieved by handling this component in a Post PCR environment. Care should also be taken to avoid cross-contamination of other samples when adding the positive control to the run. This can be avoided by sealing all other samples and negative controls before pipetting the positive control into the positive control well.

Negative control

To validate any positive findings a negative control reaction should be included every time the kit is used. For this reaction the RNase/DNase free water should be used instead of template. A negative result indicates that the reagents have not become contaminated while setting up the run. It is also known as a No Template Control or NTC.

Mycobacterium DNA is known to be highly prevalent within the air and environment generally and the negative control may therefore give a late positive signal due to environmental contamination. The interpretation of results section of this handbook gives guidance on how to interpret results where environmental contamination is evident.

Resuspension protocol

To minimize the risk of contamination with foreign DNA, we recommend that all pipetting is performed in a PCR clean environment. Ideally this would be a designated PCR lab or PCR cabinet. Filter tips are recommended for all pipetting steps.

1. Pulse-spin each tube in a centrifuge before opening.

This will ensure lyophilised primer/probe mix or template is in the base of the tube and is not lost upon opening the tube.

2. Resuspend the kit components in the RNase/DNase free water supplied, according to the table below.

To ensure complete resuspension, vortex each tube thoroughly.

Component - resuspend in water	Volume
Pre-PCR pack	
Mycobacterium spp_v2.0 primer/probe mix (BROWN)	165 µl

3. Resuspend the positive control template in the template preparation buffer supplied, according to the table below:

To ensure complete resuspension, vortex the tube thoroughly.

Component - resuspend in template preparation buffer	Volume
Post-PCR heat-sealed foil	
Mycobacterium spp_v2.0 Positive Control Template (RED)	500 µl

* This component contains high copy number template and is a VERY significant contamination risk. It must be opened and handled in a separate laboratory environment, away from the other components.

qPCR detection protocol

1. For each DNA sample prepare a reaction mix according to the table below:
Include sufficient reactions for positive and negative controls.

Component	Volume
oasig® lyophilised or PrecisionPLUS® 2X qPCR Master Mix	10 µl
Mycobacterium spp_v2.0 primer/probe mix (BROWN)	1 µl
RNase/DNase free water (WHITE)	4 µl
Final Volume	15 µl

2. Pipette 15µl of this mix into each well according to your qPCR experimental plate set up.
3. Pipette 5µl of DNA template into each well, according to your experimental plate set up.
For negative control wells use 5µl of RNase/DNase free water (**WHITE**). For positive control wells use 5 µl of the positive control template (**RED**). The final volume in each well is 20µl.

4. (Optional) Standard curve preparation for quantitative analysis.

For quantitative analysis of the samples, a standard curve dilution series can be prepared using the positive control template (**RED**). This is not required for qualitative analysis.

4.1 Reaction mix preparation for the standard curve.

Include sufficient reactions for each dilution of the standard curve.

Component	Volume
oasig [®] lyophilised or PrecisionPLUS [®] 2X qPCR Master Mix	10 µl
Mycobacterium spp_v2.0 primer/probe mix (BROWN)	1 µl
RNase/DNase free water (WHITE)	4 µl
Final Volume	15 µl

4.2 Preparation of a 10-fold standard curve dilution series.

- Pipette 90µl of template preparation buffer (**YELLOW**) into 5 tubes and label 2-6.
- Pipette 10µl of Positive Control Template (**RED**) into tube 2.
- Vortex thoroughly.
- Change pipette tip and pipette 10µl from tube 2 into tube 3
- Vortex thoroughly.

Repeat steps d and e to complete the dilution series.

Standard Curve	Copy Number
Tube 1 Positive control (RED)	2 x 10 ⁵ per µl
Tube 2	2 x 10 ⁴ per µl
Tube 3	2 x 10 ³ per µl
Tube 4	2 x 10 ² per µl
Tube 5	20 per µl
Tube 6	2 per µl

4.3 Pipette 5µl of standard template into each well for the standard curve according to your experimental plate set up.

The final volume in each well is 20µl.

qPCR Amplification Protocol

Amplification conditions using oasisig[®] lyophilised or PrecisionPLUS[®] 2X qPCR Master Mix.

	Step	Time	Temp
	Enzyme activation	2 min	95 °C
Cycling x50	Denaturation	10 s	95 °C
	DATA COLLECTION *	60 s	60 °C

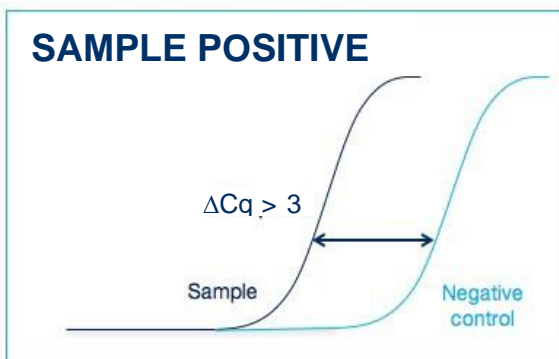
* Fluorogenic data should be collected during this step through the FAM channel.

Interpretation of results

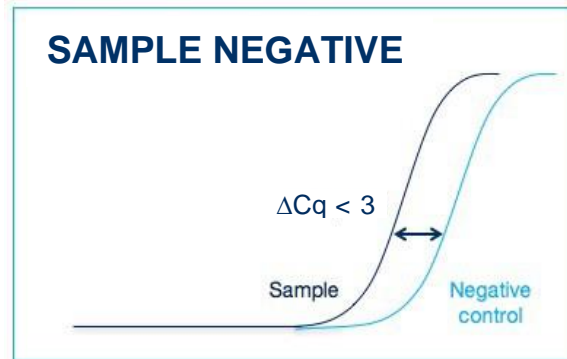
Target	Positive control	Negative control	Interpretation
+	+	-	POSITIVE QUANTITATIVE RESULT calculate copy number
-	+	-	NEGATIVE RESULT
+ / -	+	≤ 35	EXPERIMENT FAILED due to test contamination
+ / -	+	> 35	*
+ / -	-	+ / -	EXERIMENT FAILED

Positive control template is expected to amplify between Cq 16 and 23. Failure to satisfy this quality control criterion is a strong indication that the experiment has been compromised.

*Where the test sample is positive and the negative control is positive with a Cq > 35, the sample must be reinterpreted based on the relative signal strength of the two results:



If the sample amplifies > 3 Cq earlier than the negative control, then the sample should be reinterpreted (via the table above) with the negative control verified as negative.



If the sample amplifies < 3 Cq earlier than the negative control, then the positive sample result is invalidated, and a negative call is the correct result.

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