

Primerdesig™

R01054

Vibrio All Species

Kit version: 2

Target Region: rpoB gene

genesig® Standard Kit

150 tests

GENESIG

Kits by Primerdesign

Specificity of primers and probes last reviewed on:
07 July 2023

For general laboratory and research use only

Introduction to Vibrio All Species

Vibrio is a genus of “comma” shaped, facultatively anaerobic Gram-negative bacteria that are commonly present in marine environments. These bacteria are motile using a single polar flagellum, they do not form spores and can undergo both respiratory and fermentative metabolism. They are sensitive to acidic pH but can tolerate alkaline pH very well. Most species of Vibrio are halophilic and require saltwater for growth. In tropical and temperate regions, disease-causing species of Vibrio occur naturally in marine and coastal environments. They are heterotrophic organisms and derive their nutrients through a mutualistic or parasitic relationship with the host organism.

The genus contains at least twelve species that are pathogenic to humans, eight of which are associated with food-borne disease. The majority of food-borne disease is caused by *V. cholerae*, *V. parahaemolyticus* and *V. vulnificus*. Some species such as *V. cholerae* and *V. parahaemolyticus* are primarily associated with gastrointestinal disease, while others like *V. vulnificus* can cause non-intestinal illnesses, such as septicaemia. While immunosuppressed individuals are most susceptible to Vibrio infections, these bacteria are capable of being harmful to healthy individuals. Pathogenic Vibrio species, in particular *V. cholerae*, can also be recovered from freshwater reaches of estuaries, where it can be introduced by faecal contamination as well as having a natural presence in this environment.

Specificity

The Primerdesign™ genesig® Kit for bacteria species of the Vibrio genus (Vibrio spp) is designed for the in vitro quantification of Vibrio genomes. The kit is designed to have a broad detection profile. The assays within this kit are predicted to detect over 95% of sequences available on the NCBI database at the time of the most recent review.

The dynamics of genetic variation means that new sequence information may become available after the initial design. Primerdesign™ periodically reviews the detection profiles of our kits and when required releases new versions.

If you require further information or have a specific question about the detection profile of this kit, then please send an e-mail to techsupport@primerdesign.co.uk and our team will answer your question.

Kit contents

- **Vibrio_spp v2.0 primer/probe mix (150 reactions BROWN)**
FAM labelled
- **Vibrio_spp v2.0 positive control template (RED)**
- **RNase/DNase free water (WHITE)**
for resuspension of primer/probe mixes
- **2x Template preparation buffer (YELLOW)**
for resuspension of positive control template and standard curve preparation

Reagents and equipment to be supplied by the user

Real-time PCR Instrument

Extraction kit

This kit is recommended for use with genesig® Easy DNA/RNA extraction kit. However, it is designed to work well with all processes that yield high quality RNA and DNA with minimal PCR inhibitors.

Oasig™ lyophilised or Precision®PLUS 2X qPCR Master Mix

This kit is intended for use with oasig™ lyophilised or Precision®PLUS 2X qPCR Master Mix.

Pipettors and filter tips

Vortex and centrifuge

1.5 ml microtubes

qPCR plates or reaction tubes

Kit storage and stability

This kit is stable at room temperature but should be stored at -20°C on arrival. Once the lyophilised components have been resuspended, they should not be exposed to temperatures above -20°C for longer than 30 minutes at a time and unnecessary repeated freeze/thawing should be avoided. The kit is stable for six months from the date of resuspension under these circumstances.

If a standard curve dilution series is prepared this can be stored frozen for an extended period. If you see any degradation in this serial dilution a fresh standard curve can be prepared from the positive control.

Primerdesign™ does not recommend using the kit after the expiry date stated on the pack.

Suitable sample material

All kinds of sample material suited for PCR amplification can be used. Please ensure the samples are suitable in terms of purity, concentration, and DNA integrity. Always run at least one negative control with the samples. To prepare a negative control, replace the template RNA sample with RNase/DNase free water.

Dynamic range of test

Under optimal PCR conditions the kit can achieve priming efficiencies between 90-110% and detect less than 100 copies of target template. If running a positive control standard curve for a quantitative result, and an efficiency of between 90% to 110% is not achieved, then the run should be repeated with a freshly prepared standard curve.

Notices and disclaimers

This product is developed, designed and sold for research purposes only. It is not intended for human diagnostic or drug purposes or to be administered to humans unless clearly expressed for that purpose by the Food and Drug Administration in the USA or the appropriate regulatory authorities in the country of use. During the warranty period Primerdesign™ genesig® detection kits allow precise and reproducible data recovery combined with excellent sensitivity. For data obtained by violation to the general GLP guidelines and the manufacturer's recommendations the right to claim under guarantee is expired. PCR is a proprietary technology covered by several US and foreign patents. These patents are owned by Roche Molecular Systems Inc. and have been sub-licensed by PE Corporation in certain fields. Depending on your specific application you may need a license from Roche or PE to practice PCR. Additional information on purchasing licenses to practice the PCR process may be obtained by contacting the Director of Licensing at Roche Molecular Systems, 1145 Atlantic Avenue, Alameda, CA 94501 or Applied Biosystems business group of the Applied Biosystems Corporation, 850 Lincoln Centre Drive, Foster City, CA 94404. In addition, the 5' nuclease assay and other homogeneous amplification methods used in connection with the PCR process may be covered by U.S. Patents 5,210,015 and 5,487,972, owned by Roche Molecular Systems, Inc, and by U.S. Patent 5,538,848, owned by The Perkin-Elmer Corporation.

Trademarks

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Principles of the test

Real-time PCR

A *Vibrio_spp* specific primer and probe mix is provided, and this can be detected through the FAM channel.

The primer and probe mix provided exploits the so-called TaqMan® principle. During PCR amplification, forward and reverse primers hybridize to the *Vibrio_spp* DNA. A fluorogenic probe is included in the same reaction mixture which consists of a DNA probe labelled with a 5'-dye and a 3'-quencher. During PCR amplification, the probe is cleaved, and the reporter dye and quencher are separated. The resulting increase in fluorescence can be detected on a range of qPCR platforms.

Positive control

For copy number determination and as a positive control for the PCR set up, the kit contains a positive control template. This can be used to generate a standard curve of *Vibrio_spp* copy number/Cq value. Alternatively, the positive control can be used at a single dilution where full quantitative analysis of the samples is not required. Each time the kit is used, at least one positive control reaction must be included in the run. A positive result indicates that the primers and probes for detecting the target *Vibrio_spp* gene worked properly in that particular experimental scenario. If a negative result is obtained the test results are invalid and must be repeated. Care should be taken to ensure that the positive control does not contaminate any other kit component which would lead to false-positive results. This can be achieved by handling this component in a Post PCR environment. Care should also be taken to avoid cross-contamination of other samples when adding the positive control to the run. This can be avoided by sealing all other samples and negative controls before pipetting the positive control into the positive control well.

Negative control

To validate any positive findings a negative control reaction should be included every time the kit is used. For this reaction the RNase/DNase free water should be used instead of template. A negative result indicates that the reagents have not become contaminated while setting up the run. It is also known as a No Template Control or NTC.

Resuspension protocol

To minimize the risk of contamination with foreign DNA, we recommend that all pipetting be performed in a PCR clean environment. Ideally this would be a designated PCR lab or PCR cabinet. Filter tips are recommended for all pipetting steps.

1. Pulse-spin each tube in a centrifuge before opening.

This will ensure lyophilised primer and probe mix is in the base of the tube and is not lost upon opening the tube.

2. Resuspend the kit components in the RNase/DNase free water supplied, according to the table below.

To ensure complete resuspension, vortex each tube thoroughly.

Component - resuspend in water	Volume
Pre-PCR pack	
Vibrio_spp v2.0 primer/probe mix (BROWN)	165 µl

3. Resuspend the positive control template in the template preparation buffer supplied, according to the table below:

To ensure complete resuspension, vortex the tube thoroughly.

Component - resuspend in template preparation buffer	Volume
Post-PCR heat-sealed foil	
Vibrio_spp v2.0 Positive Control Template (RED) *	500 µl

* This component contains high copy number template and is a VERY significant contamination risk. It must be opened and handled in a separate laboratory environment, away from the other components.

qPCR detection protocol

1. For each DNA sample prepare a reaction mix according to the table below:
Include sufficient reactions for positive and negative controls.

Component	Volume
Oasig™ lyophilised or Precision®PLUS 2X qPCR Master Mix	10 µl
Vibrio_spp v2.0 primer/probe mix (BROWN)	1 µl
RNase/DNase free water (WHITE)	4 µl
Final Volume	15 µl

2. Pipette 15µl of this mix into each well according to your qPCR experimental plate set up.
3. Pipette 5µl of DNA template into each well, according to your experimental plate set up.
For negative control wells use 5µl of RNase/DNase free water. The final volume in each well is 20µl.
4. If a standard curve is included for quantitative analysis, prepare a reaction mix according to the table below:

Component	Volume
Oasig™ lyophilised or Precision®PLUS 2X qPCR Master Mix	10 µl
Vibrio_spp v2.0 primer/probe mix (BROWN)	1 µl
RNase/DNase free water (WHITE)	4 µl
Final Volume	15 µl

5. Preparation of a standard curve dilution series.

- Pipette 90µl of template preparation buffer into 5 tubes and label 2-6
- Pipette 10µl of Positive Control Template (**RED**) into tube 2
- Vortex thoroughly
- Change pipette tip and pipette 10µl from tube 2 into tube 3
- Vortex thoroughly

Repeat steps d and e to complete the dilution series.

Standard Curve	Copy Number
Tube 1 Positive control (RED)	2×10^5 per µl
Tube 2	2×10^4 per µl
Tube 3	2×10^3 per µl
Tube 4	2×10^2 per µl
Tube 5	20 per µl
Tube 6	2 per µl

6. Pipette 5µl of standard template into each well for the standard curve according to your experimental plate set up.

The final volume in each well is 20µl.

qPCR Amplification Protocol

Amplification conditions using Oasig™ lyophilised or Precision®PLUS 2X qPCR Master Mix.

	Step	Time	Temp
	Enzyme activation	2 min	95 °C
Cycling x50	Denaturation	10 s	95 °C
	DATA COLLECTION *	60 s	60 °C

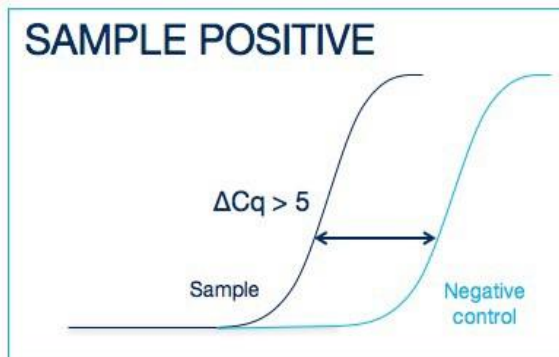
* Fluorogenic data should be collected during this step through the FAM channel

Interpretation of results

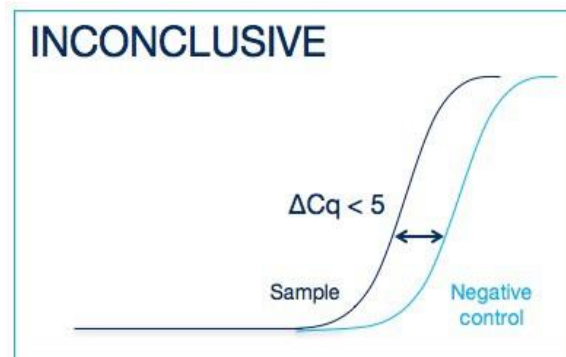
Target	Positive control	Negative control	Interpretation
+	+	-	POSITIVE QUANTITATIVE RESULT calculate copy number
-	+	-	NEGATIVE RESULT
+ / -	+	≤ 35	EXPERIMENT FAILED due to test contamination
+ / -	+	> 35	*
+ / -	-	+ / -	EXPERIMENT FAILED

Positive control template (**RED**) is expected to amplify between Cq 16 and 23. Failure to satisfy this quality control criterion is a strong indication that the experiment has been compromised.

*Where the test sample is positive and the negative control is positive with a Cq > 35 , the sample must be reinterpreted based on the relative signal strength of the two results:



If the sample amplifies > 5 Cq earlier than the negative control, then the sample should be reinterpreted (via the table above) with the negative control verified as negative.



If the sample amplifies < 5 Cq earlier than the negative control, then the positive sample result is invalidated, and the result should be determined inconclusive due to test contamination. The test for this sample should be repeated.